# Human Gonadotropin-Releasing Hormone Receptor Cell Line

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## I. Introduction

The gonadotropin-releasing hormone receptor (GnRHR) is a member of the seven-transmembrane G-protein coupled receptor (GPCR) family. It is expressed on the surface of pituitary gonadotrope cells as well as in the lymphocytes, breast, ovaries, and prostate. Following binding of gonadotropin-releasing hormone, the receptor associates with G-proteins that activate a phosphatidylinositol-calcium second messenger system. Activation of the receptor ultimately causes the release of gonadotropic luteinizing hormone (LH) and follicle-stimulating hormone (FSH). Defects in this gene are a cause of hypogonadotropic hypogonadism (HH). This manual describes establishment of a cell line and a protocol of pharmacologically validated human gonadotropin-releasing hormone receptor (Genebank Accession Number: NM\_000316).

## **II. Cell Line Information**

- Catalog Number: M00136
- > Cell Line Name: HEK/GnRHR/NFAT/βla
- > Description:

The HUMAN GnRHR is amplified by PCR using a high fidelity enzyme and subcloned into the pcDNA3.1(+) mammalian expression vector. The full-length ORF has been confirmed by sequencing. The GnRHR reporter cell line is created by cotransfection of pcDNA3.1(+)/GNRHR and pNFAT- $\beta$ la in an Hek293 cell line. The transfected cells are stably selected by 700 µg/ml G418. Single cell clones with high GnRHR inducibility and low  $\beta$ -lactamase background are isolated using ring cloning. The clones with largest dynamic ranges in  $\beta$ -lactamase are chosen for pharmacological and stability studies.

- > Function: Cell based, functional assay for GnRH receptor
- Quantity: 2 vial (2 × 10<sup>6</sup>) frozen cells
- Passage Number Shipped: 2
- Host Cell: HEK293
- Cell Phenotype: Adherent/epithelial
- Mycoplasma: Negative
- Recommended Storage: Liquid nitrogen upon delivery
- Propagation Medium: DMEM, 10% FBS, 700 µg/ml G418



### III. GnRH Dose Response of HEK/GnRHR/NFAT/βla Cell Line and Assay Procedure





- Seed 25,000 cells per well in growth medium (100 µl per well) into 96-well tissue culture treated blackwall, clear-bottom plates (Costar #3603) after trypsinization. Prepare some wells with medium alone (no cells) to use for determining plate background.
- 2. Culture cells in 5% CO<sub>2</sub> at 37°C. Allow cells to reach  $\approx$ 90% confluence.
- 3. 12-24 hours before the assay, replace Growth Medium with 100  $\mu$ I/well serum-free DMEM. Be careful not to disturb the cells.
- 4. Prepare ligand solution in serum-free DMEM (10X).
- 5. Add 10 µl of 10X ligand solution to wells for stimulation and 10 µl of serum-free DMEM per well for nonstimulated control.
- 6. Incubate cells in 5%  $CO_2$  at 37°C for 5-6 hours.
- 7. Load cells with 2  $\mu$ M CCF4/AM as described in CCF4 Loading Protocol.
- 8. Incubate the plate at room temperature for 60-120 minutes without shaking.
- 9. Read with Analyst HT plate reader or similar fluorescence reader.

### **IV. References**

- 1. Chi Keung Cheng *et al.* Molecular Biology of Gonadotropin-Releasing Hormone (GnRH)-I, GnRH-II, and Their Receptors in Humans. Endocrine Reviews 26: 283–306, 2005
- 2. JIMMY D. NEILL *et al.* Minireview: GnRH and GnRH Receptor Genes in the Human Genome. Endocrinology 143: 737–743, 2002
- 3. Karen L Herbst, Gonadotropin-releasing hormone antagonists. Current Opinion in Pharmacology 3:660–666, 2003



### V. Appendix

#### **Cell Culture Conditions**

#### **Complete Culture Medium:**

DMEM: 90%, FBS: 10%, L-glutamine 2.0 mM, Amp 100 µg/ml, Strep 100 µg/ml, G418 700 µg/ml

#### Serum-free DMEM:

Same as above but with no FBS, 0.1% BSA

#### Freezing Medium:

Complete culture medium plus 20% FBS and 10% DMSO

#### **Thawing Cells:**

- 1. Quickly thaw frozen cells in a 37°C water bath, agitating continuously.
- Using a 1 ml pipette, slowly pipet the cells up and down five times and add, drop by drop, to a 15 ml centrifuge tube containing 5 ml of fresh prewarmed complete DMEM medium. Then centrifuge at 1,000 rpm for five minutes.
- 3. Discard the supernatant medium and resuspend the cell pellet in 5 ml of fresh prewarmed complete DMEM medium. Transfer cells to a T25 flask and incubate at 37°C with 5% CO<sub>2</sub> until the cells reach >90% confluence. The recovery rate for frozen cells is usually 90% or above.

#### Subculturing:

When the cells reach confluence, they need split. This cell line is normally split twice weekly at 1:8 to 1:15 dilutions.

- 1. Carefully aspirate all the media. Gently rinse the cell layer with appropriate amount of 0.2% trypsin-EDTA, and aspirate it off.
- 2. Wait for about 1-3 minutes. Dislodge the cells by gently tapping the sides of flask or dish.
- 3. Resuspend cells with appropriate amount of complete DMEM medium, and split cells as desired.

#### **Changing Medium:**

This is normally done every other day.

- 1. Gently aspirate off medium.
- 2. Transfer fresh warm complete DMEM medium (37°C) into a flask (5 ml for T25 and 10 ml for T75).



#### **Freezing Cells:**

- 1. Repeat steps 1-3 of subculturing section.
- 2. Centrifuge down the cells at 1,000 rpm for five minutes.
- Aspirate off the supernatant and resuspend the cells in fresh freezing medium at a density of 2-3 x 10<sup>6</sup> cells/ml. Add 1 ml cells per cryogenic vial.
- 4. Put the cryogenic vial of cells into cryo freezing container. Then transfer the container to a -80°C environment and leave it there overnight.
- 5. Transfer cryogenic vial into liquid nitrogen (-196°C).

#### **Reagents & Consumables:**

- 1. DMEM: Dulbecco's Modified Eagle Medium powder, high glucose (Gibco BRL, Cat #12100-046)
- 2. FBS: Fetal Bovine Serum (Hyclone, Cat #CH30160.03)
- 3. L-Glutamine: 200 mM (Gibco BRL, Cat # 25030-081)
- 4. Ampicillin: 50 mg/ml (Sigma A-9518)
- 5. Streptomycin Sulfate: 50 mg/ml (Gibco BRL, Cat # 11860-038)
- 6. Hygromycin B in PBS, 50 mg/ml (Invitrogen, Cat #10687-010)
- 7. Trypsin: 1:250 rom Bovine Pancreas (Gibco BRL, Cat # 27250016)
- 8. DMSO: dimethyl sulphoxide (Sigma, Cat #D8418)
- 9. Hepes: Sigma Cat #H-3375
- 10. CCF4: (Invitrogen, Cat #K1096)
- 11. GnRH: synthesized
- 12. Venor<sup>®</sup>GeM Mycoplasma Detection kit: Minerva Biolabs Cat #11-1050
- 13. 96 Well Plate: Costar, Cat# 3603, Blackwall/clear bottom, Polystyrene, sterilized.

#### Media and Solutions:

1. PBS (for preparation of 500 ml)

1)	KCI:	0.1 g
2)	KH <sub>2</sub> PO <sub>4</sub> :	0.1 g
3)	NaCI:	4.0 g
4)	Na <sub>2</sub> HPO <sub>4</sub> .12H <sub>2</sub> O:	1.4425 g

Dissolve the above components in double-distilled water (ddH<sub>2</sub>O) and adjust pH to 7.4 with 0.1 N NaOH. Add ddH<sub>2</sub>O to the final volume of 500 ml. Autoclave and store at 4°C.

2. Trypsin-EDTA (for preparation of 100 ml)

1)	Trypsin:	•	0.25 g
2)	2%EDTA:		2 ml
3)	PBS:		98 ml

Dissolve trypsin in 2%EDTA and PBS completely; sterilize the solution by passing through a 0.20  $\mu m$  membrane filter; store at 4°C.



- 3. Culture medium (for preparation of 1 L)
  - 1) Measure out 950 ml distilled water to dissolve the media components with gentle stirring until the solution becomes clear.
  - 2) Add NaHCO<sub>3</sub> 3.7 g for high glucose DMEM
  - Adjust pH of medium to 0.2-0.3 below the desired final working pH (using 1 N NaOH or 1 N HCL is recommended). Add slowly with stirring.
  - 4) Dilute to 1 liter with  $ddH_2O$ .
  - 5) Sterilize the medium immediately using the method of membrane filtration. Store at 4°C
- 4. Ampicillin/Streptomycin 50 mg/ml

Dissolve 1 g Ampicillin or Streptomycin in 20 ml ddH<sub>2</sub>O and sterilize the solution by membrane filtration using 0.20  $\mu$ m filter. Aliquot and store at 4°C for short-term conservation and -20°C for long term conservation.

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